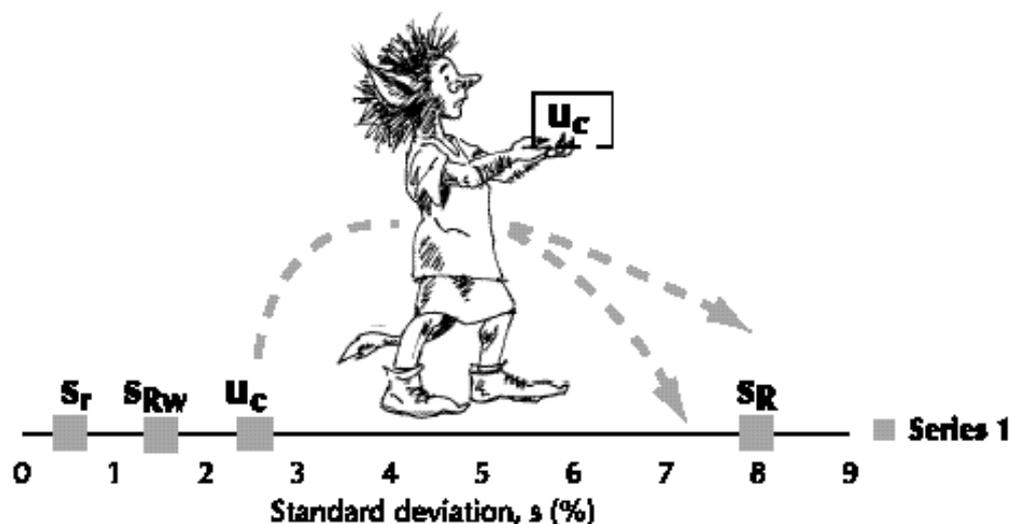


# HANDBOOK FOR CALCULATION OF MEASUREMENT UNCERTAINTY IN ENVIRONMENTAL LABORATORIES



**Bertil Magnusson**  
**Teemu Näykki**  
**Håvard Hovind**  
**Mikael Krysell**



<p><b>Authors:</b> Bertil Magnusson<sup>1</sup> Teemu Näykki<sup>2</sup> Håvard Hovind<sup>3</sup> Mikael Krysell<sup>4</sup></p>	<p><b>NORDTEST project number:</b> 1589-02</p>	
	<p><b>Institution:</b> <sup>1)</sup> SP, Sweden, <sup>2)</sup> SYKE, Finland, <sup>3)</sup> NIVA, Norway, <sup>4)</sup> Eurofins A/S, Denmark</p>	
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<p><b>Title (Original):</b></p>	<p>Handbook for Calculation of Measurement Uncertainty in Environmental Laboratories</p>	
<p><b>Abstract:</b></p> <p>This handbook is written for environmental testing laboratories in the Nordic countries, in order to give support to the implementation of the concept of measurement uncertainty for their routine measurements. The aim is to provide a practical, understandable and common way of measurement uncertainty calculations, mainly based on already existing quality control and validation data, according to the European accreditation guideline /12/, the Eurolab Technical Report No. 1 /3/ and the ISO/DTS 21748 Guide /8/. Nordtest has supported this project economically in order to promote and enhance harmonisation between laboratories on the Nordic market.</p> <p>Practical examples, taken directly from the everyday world of environmental laboratories, are presented and explained. However, the approach is very general and should be applicable to most testing laboratories in the chemical field.</p> <p>The handbook covers all steps in the analytical chain from the arrival of the sample in the laboratory until the data has been reported. It is important to notice that vital parts of the total measurement uncertainty are not included, e.g. sampling, sample transportation and possible gross errors during data storage/retrieval.</p> <p>The recommendations in this document are primarily for guidance. It is recognised that while the recommendations presented do form a valid approach to the evaluation of measurement uncertainty for many purposes, other suitable approaches may also be adopted – see references in Section 9. Especially the EURACHEM/CITAC-Guide /2/ is useful in cases where sufficient previous data is not available, and therefore the mathematical analytical approach according to GUM /1/ with all different steps is to be used.</p> <p>Basic knowledge in the use of quality control and statistics is required. In order to make it possible for the reader to follow the calculations, some raw data is given in appendices.</p>		
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for  
Calculation of  
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**Project participants**

Bertil Magnusson, SP, Sweden  
Teemu Näykki, SYKE, Finland  
Håvard Hovind, NIVA, Norway  
Mikael Krysell, Eurofins A/S, Denmark

Drawings by Petter Wang, NIVA, Norway

**Valuable comments on the  
contents have been provided by:**

Rolf Flykt, Sweden  
Irma Mäkinen, Finland  
Ulla O. Lund, Denmark  
Steve Ellison, UK



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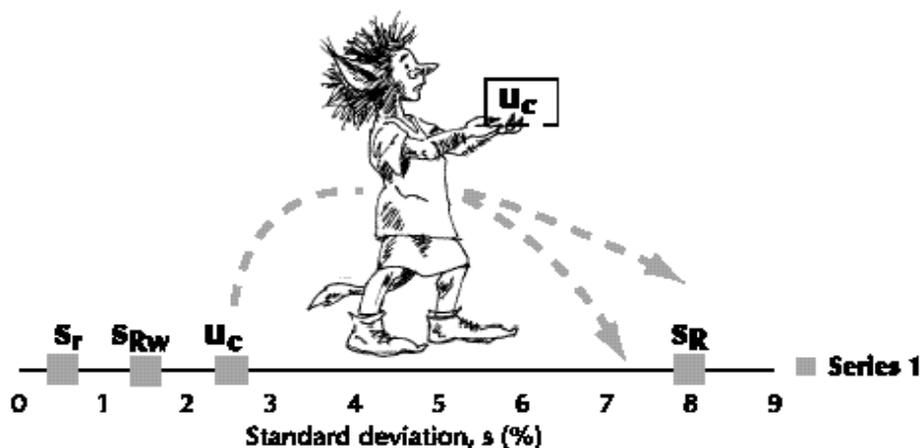


# 1 Definitions and abbreviations

$s$	An estimate of the population standard deviation $\sigma$ from a limited number (n) of observations ( $x_i$ )
$\bar{x}$	Mean value
$u(x)$	Individual standard uncertainty component (GUM, /1/).
$u_c$	Combined standard uncertainty (GUM, /1/)
$U$	Expanded combined uncertainty close to 95 % confidence interval
$r$	<p>Repeatability limit – performance measure for a test method or a defined procedure when the test results are obtained under <b>repeatability conditions</b>.</p> <p><b>Repeatability conditions:</b> Conditions where independent test results are obtained with the same method on identical test items in the same laboratory by the same operator using the same equipment within short intervals of time.</p> <p>Repeatability (precision under repeatability conditions) is also sometimes called “within run precision” (ISO 3534-1, /6/).</p>
$s_r$	Repeatability standard deviation of a measurement (can be estimated from a series of duplicate analyses)
$R$	<p>Reproducibility limit – performance measure for a test method or procedure when the test results are obtained under <b>reproducibility conditions</b>.</p> <p><b>Reproducibility conditions:</b> Conditions where test results are obtained with the same method on identical test items in different laboratories with different operators using different equipment.</p> <p>Reproducibility (precision under reproducibility conditions) is also sometimes called “between lab precision” (ISO 3534-1, /6/).</p>
$s_R$	<p>Reproducibility standard deviation of a measurement (can be estimated from validation studies with many participating laboratories or from other interlaboratory comparisons e.g. proficiency testing data)</p> <p>Note: <math>R = 2.8 \cdot s_R</math></p>
$R_w$	Within-laboratory reproducibility = intermediate measure between $r$ and $R$ , where <i>operator</i> and/or <i>equipment</i> and/or <i>time</i> and/or <i>calibration</i> can be varied, but in the same laboratory. An alternative name is intermediate precision
$s_{Rw}$	Reproducibility within-laboratory standard deviation (can be estimated from standard deviation of a control sample over a certain period of time, preferably one year)

<i>CRM</i>	Certified Reference Material
<i>Certified value</i>	Assigned value given to a CRM, quantified through a certification process (traceable to SI-unit and with a known uncertainty)
<i>Nominal value</i>	Nominal value is the assigned value, e.g. in an interlaboratory comparison where it is the organiser's best representation of the "true value"
$u(Cref)$	Uncertainty component from the certified or nominal value
<i>bias</i>	Difference between mean measured value from a large series of test results and an accepted reference value (a certified or nominal value). The measure of trueness is normally expressed in term of bias. Bias for a measurement, e.g. for a laboratory or for an analytical method..
$u(bias)$	Uncertainty component for bias. The $u(bias)$ , is always included in the measurement uncertainty calculations
$RMS_{bias}$	$\sqrt{\frac{\sum (bias_i)^2}{n}}$
<i>Interlaboratory comparison</i>	General term for a collaborative study for either method performance, laboratory performance (proficiency testing) or material certification.

- Ammonium-values for repeatability**  $s_r$
- Reproducibility within laboratory**  $s_{Rw}$
- Combined uncertainty**  $U_c$
- Reproducibility between laboratories**  $s_R$



## 2 Introduction

### 2.1 *Scope and field of application*

This handbook is written for environmental testing laboratories in the Nordic countries, in order to give support to the implementation of the concept of measurement uncertainty for their routine measurements. The aim is to provide a practical, understandable and common way of measurement uncertainty calculations, mainly based on already existing quality control and validation data, according to the European accreditation guideline /12/, the Eurolab Technical Report No. 1 /3/ and the ISO/DTS 21748 Guide /8/. Nordtest has supported this project economically in order to promote and enhance harmonisation between laboratories on the Nordic market.

Practical examples, taken directly from the everyday world of environmental laboratories, are presented and explained. However, the approach is very general and should be applicable to most testing laboratories in the chemical field.

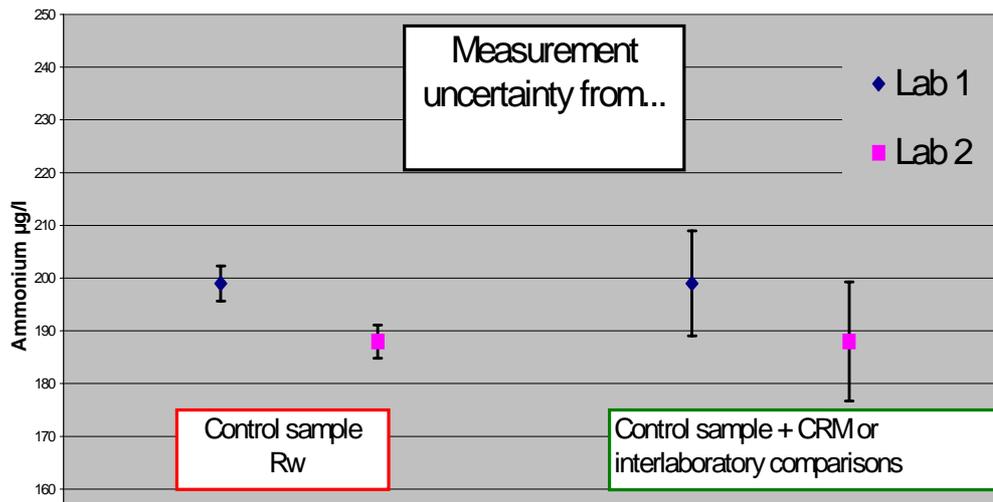
The handbook covers all steps in the analytical chain from the arrival of the sample in the laboratory until the data has been reported. It is important to notice that vital parts of the total measurement uncertainty are not included, e.g. sampling, sample transportation and possible gross errors during data storage/retrieval.

The recommendations in this document are primarily for guidance. It is recognised that while the recommendations presented do form a valid approach to the evaluation of measurement uncertainty for many purposes, other suitable approaches may also be adopted – see references in Section 9. Especially the EURACHEM/CITAC-Guide /2/ is useful in cases where sufficient previous data is not available, and therefore the mathematical analytical approach according to GUM /1/ with all different steps is to be used.

Basic knowledge in the use of quality control and statistics is required. In order to make it possible for the reader to follow the calculations, some raw data is given in appendices

### 2.2 *Comment to customers*

Previously, laboratories usually reported uncertainty as the standard deviation calculated from data for an internal control sample. The measurement uncertainty also taking into account method and laboratory bias and using a coverage factor of 2, can give uncertainty values which may be a factor of 2 to 5 times higher than previously (Figure 1). However, this does not reflect a change in the performance of the laboratory, just a much better estimation of the real variation between laboratories. In Figure 1, the ammonium results from two laboratories are in good agreement – the difference is about 5 %. You can see this if you look to the right where measurement uncertainty is calculated correctly, but not if you look to the left, where the uncertainty is calculated directly from a control sample and presented as the standard deviation ( $\pm 1s$ ).



**Figure 1.** Comparing ammonium results from two laboratories, Lab 1 = 199 µg/L and Lab 2 = 188 µg/L. To the left the error bars are calculated from results on control samples ( $\pm 1s$ ) and to the right the error bars are expanded measurement uncertainty.

### 2.3 About Measurement Uncertainty

What is measurement uncertainty?

- The number after  $\pm$
- All measurements are affected by a certain error. The measurement uncertainty tells us what size the measurement error **might** be. Therefore, the measurement uncertainty is an important part of the reported result
- Definition: Measurement uncertainty is "A parameter associated with the result of a measurement, that characterises the dispersion of the values that could reasonably be attributed to the measurand" /1, 5/

Who needs measurement uncertainties?

- The customer needs it together with the result to make a correct decision. The uncertainty of the result is important, e.g. when looking at allowable (legal) concentration limits
- The laboratory to know its own quality of measurement and to improve to the required quality

Why should the laboratory give measurement uncertainty?

- As explained above, the customers need it to make correct decisions
- An estimation of the measurement uncertainty is required in ISO 17025 /9/

#### How is measurement uncertainty obtained?

- The basis for the evaluation is a measurement and statistical approach, where the different uncertainty sources are estimated and combined into a single value
- “*Basis for the estimation of measurement uncertainty is the existing knowledge (no special scientific research should be required from the laboratories). Existing experimental data should be used (quality control charts, validation, interlaboratory comparisons, CRM etc.)*” /12/
- Guidelines are given in GUM /1/, further developed in, e.g., EA guidelines /12/, the Eurachem/Citac guide /2/, in a Eurolab technical report /3/ and in ISO/DTS 21748 /8/

#### How is the result expressed with measurement uncertainty?

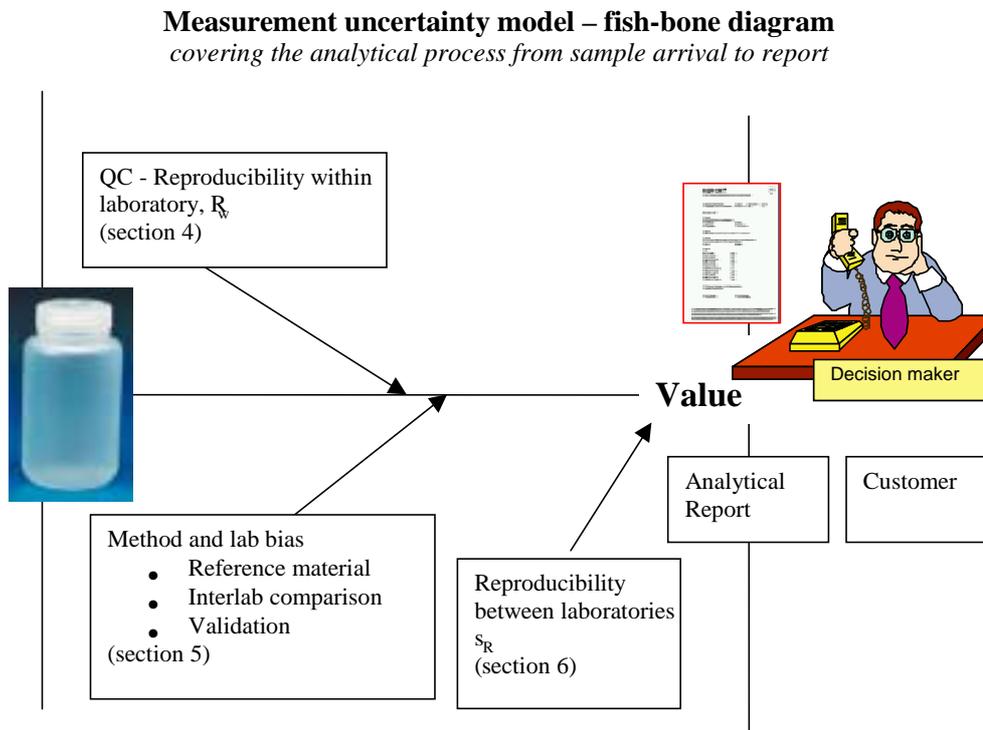
- Measurement uncertainty should normally be expressed as  $U$ , the combined expanded measurement uncertainty, using a coverage factor  $k = 2$ , providing a level of confidence of approximately 95 %
- It is often useful to state how the measurement uncertainty was obtained
- Example, where  $\pm 7$  is the measurement uncertainty:  
Ammonium ( $\text{NH}_4\text{-N}$ ) =  $148 \pm 7 \mu\text{g/L}$ . The measurement uncertainty,  $7 \mu\text{g/L}$  (95 % confidence level, i.e. the coverage factor  $k=2$ ) is estimated from control samples and from regular interlaboratory comparisons

#### How should measurement uncertainty be used?

- It can be used as in Figure 1, to decide whether there is a difference between results from different laboratories, or results from the same laboratory at different occasions (time trends etc.)
- It is necessary when comparing results to allowable values, e.g. tolerance limits or allowable (legal) concentrations

### 3 Calculation of expanded uncertainty, U - overview

A common way of presenting the different contributions to the total measurement uncertainty is to use a so-called fish-bone (or cause-and-effect) diagram. We propose a model (Figure 2), where either the reproducibility within-laboratory ( $R_w$ ) is combined with estimates of the method and laboratory bias, (error model in Appendix 3) or the reproducibility  $s_R$  is used more or less directly, ISO Guide 21748/8/. The alternative way is to construct a detailed fish-bone diagram and calculate/estimate the individual uncertainty contributions. This approach may prove very useful when studying or quantifying individual uncertainty components. It has been shown, though, that in some cases this methodology underestimates the measurement uncertainty /3/, partly because it is hard to include all possible uncertainty contributions in such an approach. By using existing and experimentally determined quality control (QC) and method validation data, the probability of including all uncertainty contributions will be maximised.



**Figure 2.** Measurement uncertainty model (fish-bone diagram), where the reproducibility within-laboratory is combined with estimates of the method and laboratory bias. Alternatively, according to ISO guide 21748 /8/, the combined uncertainty  $u_c$  can be directly estimated from the reproducibility between laboratories ( $s_R$ ). This approach is treated in section 6

### **3.1 Customer needs**

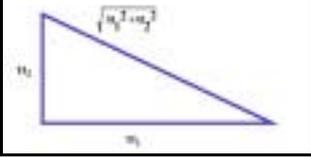
Before calculating or estimating the measurement uncertainty, it is recommended to find out what are the needs of the customers. After that, the main aim of the actual uncertainty calculations will be to find out if the laboratory can fulfil the customer demands with the analytical method in question. However, customers are not used to specifying demands, so in many cases the demands have to be set in dialogue with the customer. In cases where no demands have been established, a guiding principle could be that the calculated expanded uncertainty,  $U$ , should be approximately equal to, or less than, 2 times the reproducibility,  $s_R$ .

### **3.2 Flow scheme for uncertainty calculations**

The flow scheme presented in this section forms the basis for the method outlined in this handbook. The flow scheme, involving 6 defined steps, should be followed in all cases. The example with  $\text{NH}_4\text{-N}$  in water shows the way forward for calculating the measurement uncertainty using the flow scheme. Explanations of the steps and their components will follow in the succeeding chapters. For each step, there may be one or several options for finding the desired information.

*Background for the  $\text{NH}_4\text{-N}$  example – automatic photometric method:* The laboratory has participated in 6 interlaboratory comparisons recently. All results have been somewhat higher than the nominal value. The laboratory therefore concludes that there may be a small positive bias. On average, the bias has been +2.2 %. This bias is considered small by the laboratory and is not corrected for in their analytical results, but exists, and is thus another uncertainty component.

For this method, the main sources of uncertainty are contamination and variation in sample handling, both causing random uncertainty components. These uncertainty sources will be included in the calculations below.

Step	Action	Example – Ammonium $\text{NH}_4\text{-N}$
1	<b>Specify Measurand</b>	Ammonium is measured in water according to EN/ISO 11732 /11/. The customer demand on expanded uncertainty is $\pm 10 \%$
2	<b>Quantify <math>R_w</math> comp. A control sample B possible steps not covered by the control sample</b>	A: Control limits are set to $\pm 3.34 \%$ (95 % confidence limit) B: The control sample includes all analytical steps.
3	<b>Quantify bias comp.</b>	From interlaboratory comparisons over the last 3 years the bias result were 2.4; 2.7; 1.9; 1.4; 1.8; and 2.9. The root mean square (RMS) of the bias is 2.25 %. The uncertainty of the nominal values is $u(\text{Cref}) = 1.5 \%$ . (see Appendix 4 for explanations)
4	<b>Convert components to standard uncertainty <math>u(x)</math></b>	Confidence intervals and similar distributions can be converted to standard uncertainty /1, 2, 14/. $u(R_w) = 3.34/2 = 1.67 \%$ $u(\text{bias}) = \sqrt{\text{RMS}_{\text{bias}}^2 + u(\text{Cref})^2}$ $= \sqrt{2.25^2 + 1.5^2} = 2.71 \%$
5	<b>Calculate combined standard uncertainty, <math>u_c</math></b> 	Standard uncertainties can be summed by taking the square root of the sum of the squares $u_c = \sqrt{u(R_w)^2 + (u(\text{bias}))^2} = \sqrt{1.67^2 + 2.71^2} = 3.18$
6	<b>Calculate expanded uncertainty, <math>U = 2 \cdot u_c</math></b>	The reason for calculating the expanded uncertainty is to reach a high enough confidence (app. 95 %) in that the “true value” lies within the interval given by the measurement result $\pm$ the uncertainty. $U = 2 \cdot 3.18 = 6.36 \approx 6 \%$ .

The measurement uncertainty for  $\text{NH}_4\text{-N}$  will thus be reported as  $\pm 6 \%$  at this concentration level.

### 3.3 Summary table for uncertainty calculations

The results of the calculations done in the flow scheme will then be summarised in a summary table.

#### Ammonium in water by EN/ISO 11732

Measurement uncertainty  $U$  (95 % confidence interval) is estimated to  $\pm 6\%$ . The customer demand is  $\pm 10\%$ . The calculations are based on control chart limits and interlaboratory comparisons.

		<i>Value</i>	<i>Relative <math>u(x)</math></i>	<i>Comments</i>
<b>Reproducibility within-laboratory, <math>R_w</math></b>				
Control sample $\bar{X} = 200 \mu\text{g/L}$	$R_w$	Control limits is set to $\pm 3.34\%$	1.67 %	
Other components		--		
<b>Method and laboratory bias</b>				
Reference material	bias	--		
Interlaboratory comparisons	bias	$RMS_{bias} = 2.25\%$ $u(C_{ref}) = 1.5\%$	2.71 %	$u(bias) = \sqrt{RMS_{bias}^2 + u(C_{ref})^2}$
Recovery test	bias	--		
<b>Reproducibility between laboratories</b>				
Interlaboratory comparisons	$R$	--	8.8 %	Data - see Section 6.2
Standard method	$R$	--		

Combined uncertainty,  $u_c$  is calculated from the control sample limits and bias estimation from interlaboratory comparisons. The  $s_R$  from interlaboratory comparisons can also be used (see 6.2) if a higher uncertainty estimation is acceptable.

<i>Measurand</i>	<i>Combined Uncertainty <math>u_c</math></i>	<i>Expanded Uncertainty <math>U</math></i>
Ammonium	$\sqrt{1.67^2 + 2.67^2} = 3.18\%$	$3.18 \cdot 2 = 6.4 \approx 6\%$

## 4 Reproducibility within-laboratory - $u(R_w)$

In this section the most common ways of estimating the reproducibility within-laboratory component,  $u(R_w)$ , for the measurement uncertainty calculation are explained:

- Stable control samples covering the whole analytical process. Normally one sample at low concentration level and one at a high concentration level.
- Control samples not covering the whole analytical process. Uncertainties estimated from control samples and from duplicate analyses of real samples with varying concentration levels.
- Unstable control samples.

It is of utmost importance that the estimation must cover all steps in the analytical chain and all types of matrices – worst-case scenario. The control sample data should be run in exactly the same way as the samples e.g. if the mean of duplicate samples is used for ordinary samples, then the mean of duplicate control samples should be used for the calculations.

It is likewise important to cover long-term variations of some systematic uncertainty components **within** the laboratory, e.g. caused by different stock solutions, new batches of critical reagents, recalibrations of equipment, etc. In order to have a representative basis for the uncertainty calculations and to reflect any such variation the number of results should ideally be more than 50 and cover a time period of approximately one year, but the need differs from method to method.

### 4.1 *Customer demands*

Some laboratories choose to use the customer demand when setting the limits in their control charts. The actual performance of the method is not interesting, as long as it meets the customer demands on expanded uncertainty. If, for example, the customer asks for data with an (expanded) measurement uncertainty of  $\pm 10\%$ , then, from our experience, a good starting point is to set the control limits  $\pm 5\%$ . The  $u(R_w)$  used in the calculations will then be  $2.5\%$ .<sup>1</sup> This is just a proposal and the measurement uncertainty calculations will show if these control limits are appropriate.

### 4.2 *Control sample covering the whole analytical process*

When a stable control sample is covering the whole analytical process and has a matrix similar to the samples, the within-laboratory reproducibility at that concentration level can simply be estimated from the analyses of the control

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<sup>1</sup> Treating the control limits according to GUM /1/ as type B estimate with 95 % confidence limit

samples. If the analyses performed cover a wide range of concentration levels, also control samples of other concentration levels should be used. Example: For NH<sub>4</sub>-N two control sample levels (20 µg/L and 250 µg/L) were used during year 2002. The results for the manual analysis method are presented in the table below.

		<i>Value</i>	<i>Relative u(x)</i>	<i>Comments</i>
<b>Reproducibility within-laboratory, <math>R_w</math></b>				
Control sample 1 $\bar{X} = 20.01 \mu\text{g/L}$	$s_{Rw}$	Standard deviation 0.5 µg/L	2.5 %	From measurements in 2002, n = 75
Control sample 2 $\bar{X} = 250.3 \mu\text{g/L}$	$s_{Rw}$	Standard deviation 3.7 µg/L	1.5 %	From measurements in 2002, n = 50
Other components		--		

### 4.3 Control sample for different matrices and concentration levels

When a synthetic control solution is used for quality control, and the matrix type of the control sample is not similar to the natural samples, we have to take into consideration uncertainties arising from different matrices. Example: To estimate the matrix based uncertainties, duplicate analysis of ammonium is performed, and the  $s_r$  is estimated from the corresponding R%-chart (Range%-chart /13/), giving the repeatability of analysing natural samples having a normal matrix variation at different concentration levels.

The data set consists of 73 duplicate analyses in the range of 2 µg/L – 16000 µg/L. Most of the results were below 200 µg/L. The data is divided into two parts:

$$< 15 \mu\text{g/L} \text{ and } > 15 \mu\text{g/L}$$

The  $s_r$  can be estimated from R%-charts constructed for both concentration ranges. The data is given in Appendix 5. The standard deviation is estimated from the range (see Appendix 8):  $s = \text{range} / 1.128$ .

		<i>Value</i>	<i>Relative u(x)</i>	<i>Comments</i>
<b>Reproducibility within-laboratory, <math>R_w</math></b>				
Variation from duplicate analysis 2-15 µg/L: > 15 µg/L:	$s_R$		5.7 % 3.6 %	n = 43 ( $\bar{X} = 6.50 \mu\text{g/L}$ ) n = 30 ( $\bar{X} = 816 \mu\text{g/L}$ )

At low levels it is often better to use an absolute uncertainty rather than a relative, as relative numbers tend to become extreme at very low concentrations. In this example the absolute  $u(r)$  becomes 0.37  $\mu\text{g/L}$  for the natural sample (mean concentration 7  $\mu\text{g/L}$ ) and 0.5  $\mu\text{g/L}$  for the control sample in Section 4.2 (mean concentration 20  $\mu\text{g/L}$ ).

As the estimate from duplicate analysis gives the repeatability component ( $s_r$ ) only, it should be combined with the control sample results from Section 4.2 to give a better estimate of  $s_{Rw}$ . This way, the repeatability component will be included two times, but it is normally small in comparison to the between-days variation.

		<i>Value</i>	<i>u(x)</i>	<i>Comments</i>
<b>Reproducibility within-laboratory, <math>R_w</math></b>				
Low level (2-15 $\mu\text{g/L}$ )	$s_{Rw}$	0.5 $\mu\text{g/L}$ from control sample and 0.37 $\mu\text{g/L}$ from duplicates	0.6 $\mu\text{g/L}$	Absolute $u(x) = \sqrt{0.5^2 + 0.37^2}$
High level (> 15 $\mu\text{g/L}$ )	$s_{Rw}$	1.5% from control sample and 3.6% from duplicates	3.9 %	Relative $u(x) = \sqrt{1.5^2 + 3.6^2}$

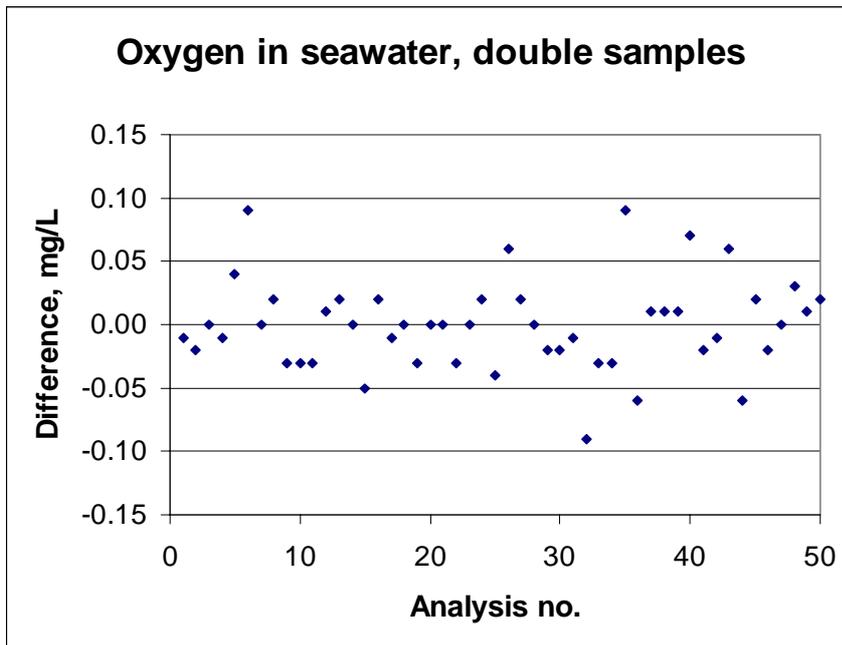
It can be noticed that the sample matrix has some effect on the variation of the results. The reason for this is not only the matrix, but also the concentration level, as almost all of the duplicate analyses were performed at a concentration level below 10  $\mu\text{g/L}$ . The quantification limit of the measurement was 2  $\mu\text{g/L}$  and the relative standard deviation usually becomes higher near that limit (cf. Figures 4 and 5 in Section 6.3).

#### 4.4 Unstable control samples

If the laboratory does not have access to stable control samples, the reproducibility can be estimated using analysis of natural duplicate samples. The results from the duplicate sample analysis can either be treated in an R-chart, where the difference between the first and second analysis is plotted directly, or as an R %-chart, where the absolute difference between the sample pair is calculated in % of the average value of the sample pair. The latter approach is particularly useful when the concentration varies a lot from time to time.

In this example, duplicate samples for oxygen have been analysed on 50 occasions. The raw data is given in Appendix 6. The concentration variation is limited, so an R-chart approach is chosen. The difference between the first and the second analysis is calculated and plotted in a chart, see Figure 3. In this case, the second result is always subtracted from the first when constructing the R-chart, as it is important to look for systematic differences between the first and the second

sample. The standard deviation for the results can be estimated from the average range of the duplicate samples (see Appendix 8), and in this case becomes 0.024. The control limits at  $\pm 2s$  thus lies at  $\pm 0.048$ . The average value of the first determination is 7.53, and the  $s_r$  thus equals  $100 \cdot 0.024 / 7.53 = 0.32\%$ .



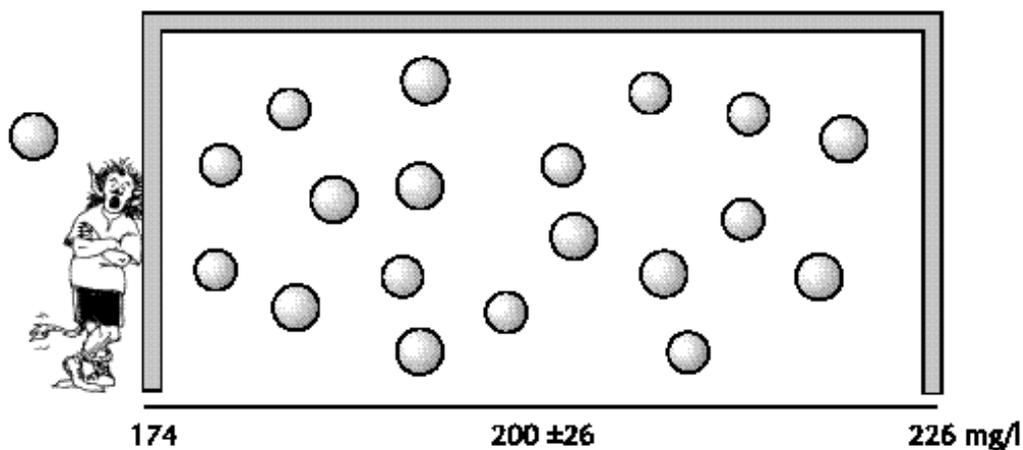
**Figure 3.** The difference between oxygen duplicate measurements plotted in an R-chart

However, this only gives the within-day variation (repeatability,  $s_r$ ) for sampling and measurement, and there will also be a “long-term” uncertainty component from the variation in the calibration (here the thiosulphate used for titrating or the calibration of the oxygen probe, depending on method). For this particular analysis, the uncertainty component from the long-term variation in calibration is hard to measure, as no stable reference material or CRM is available for dissolved oxygen. One method would be to calibrate the same thiosulphate solution several times during a few days time, and use the variation between the results. Here we choose to estimate that component by a qualified guess, but laboratories are encouraged to also try the experimental approach.

The total reproducibility within-laboratory for dissolved oxygen thus becomes:

		<i>Value</i>	<i>Relative u(x)</i>	<i>Comments</i>
<b>Reproducibility within-laboratory, R<sub>w</sub></b>				
Duplicate measurements of natural samples, difference used in r-chart	<i>s<sub>R</sub></i>	<i>s</i> = 0.024 mg/L $\bar{X}$ = 7.53 mg/L	0.32 %	Measurements in 2000-2002, n= 50
Estimated variation from differences in calibration over time		<i>s</i> = 0.5 %	0.5 %	Estimate, based on experience
<b>Combined uncertainty for R<sub>w</sub></b>				
Repeatability + reproducibility in calibration		$\sqrt{0.32^2 + 0.5^2} = 0.59 \%$		

## BOD



## 5 Method and Laboratory bias – u(bias)

In this chapter the most common ways of estimating the bias components will be outlined, namely the use of CRM, participation in interlaboratory comparisons (proficiency test) and recovery tests. Sources of bias should always be eliminated if possible. According to GUM /1/ a measurement result should always be corrected if the bias is significant and based on reliable data such as a CRM. However, even if the bias is zero, it has to be estimated and treated as an uncertainty component. In many cases the bias can vary depending on changes in matrix. This can be reflected when analysing several matrix CRMs, e.g. the bias could be both positive and negative. Examples are given and explained for the proposed calculations.

For every estimation of the uncertainty from the method and laboratory bias, two components have to be estimated to obtain  $u(bias)$ :

- 1) the bias (as % difference from the nominal or certified value)
- 2) the uncertainty of the nominal/certified value,  $u(Cref)$  or  $u(Crecovery)$

The uncertainty of the bias,  $u(bias)$  can be estimated by

$$u(bias) = \sqrt{RMS_{bias}^2 + u(Cref)^2} \text{ where } RMS_{bias} = \sqrt{\frac{\sum (bias_i)^2}{n}}$$

and if only one CRM is used also the  $s_{bias}$  have to be included and  $u(bias)$  can be estimated /14, 15/ by

$$u(bias) = \sqrt{(bias)^2 + \left(\frac{s_{bias}}{\sqrt{n}}\right)^2 + u(Cref)^2}$$

### 5.1 Certified Reference Material

Regular analysis of a CRM can be used to estimate the bias. The reference material should be analysed in at least 5 different analytical series (e.g. on 5 different days) before the values are used.

In this example the certified value is  $11.5 \pm 0.5$ , with a 95 % confidence interval.

<i>Uncertainty component from the uncertainty of the certified value</i>	
<b>Step</b>	<b>Step</b>
Convert the confidence interval to $u(Cref)$	The confidence interval is $\pm 0.5$ . Divide this by 1.96 to convert it to standard uncertainty: $0.5/1.96 = 0.26$
Convert to relative uncertainty $u(Cref)$	$100 \cdot (0.26/11.5) = 2.21\%$

**3** **Quantify Method and laboratory bias** bias =  $100 \cdot (11.9 - 11.5) / 11.5 = 3.48 \%$   
 $s_{bias} = 2.2 \%$  (n = 12)  
 $u(Cref) = 2.21 \%$

**4** **Convert components to standard uncertainty  $u(x)$**

$$u(bias) = \sqrt{(bias)^2 + \left(\frac{s_{bias}}{\sqrt{n}}\right)^2 + u(Cref)^2} =$$

$$\sqrt{(3.48)^2 + \left(\frac{2.2}{\sqrt{12}}\right)^2 + 2.21^2} = 4.2 \%$$

If **several CRM:s** are used, we will get different values for the bias. The uncertainty of the bias estimation will be calculated in the following way (see also section 5.2).

**3** **Quantify Method and laboratory bias** bias CRM1 is 3.48%, s=2,2 (n=12),  $u(Cref)=2.21 \%$   
 bias CRM2 is -0.9% s=2,0 (n=7),  $u(Cref)=1.8 \%$   
 bias CRM3 is 2.4%, s= 2,8 (n=10),  $u(Cref)=1.8 \%$   
 For the bias the  $RMS_{bias} = 2.50$   
 mean  $u(Cref)=1,9 \%$

**4** **Convert components to standard uncertainty  $u(x)$**

$$u(bias) = \sqrt{RMS_{bias}^2 + u(Cref)^2}$$

$$\sqrt{2.50^2 + 1.9^2} = 3.1 \%$$

## 5.2 Interlaboratory comparisons

In this case the results from interlaboratory comparisons are used in the same way as a reference material, i.e. to estimate the bias. In order to have a reasonably clear picture of the bias from interlaboratory comparison results, a laboratory should participate at least 6 times within a reasonable time interval.

Biases can be both positive and negative. Even if the results appear to give positive biases on certain occasions and negative on others, all bias values can be used to estimate the uncertainty component,  $RMS_{bias}$ .

The way forward is very similar to that for reference materials. However, the estimation of the bias from interlaboratory comparisons has more uncertainty to it, and thus usually becomes a bit higher than if CRMs are used. This is partly due to the fact that the certified value of a CRM normally is better defined than a nominal or assigned value in an interlaboratory comparison exercise. In some cases the calculated uncertainty  $u(Cref)$  from an interlaboratory comparison becomes too high and is not valid for estimating the  $u(bias)$ .

<i>Uncertainty component from the uncertainty of the nominal value</i>	
<b>Step</b>	<b>Example</b>
Find the between laboratory standard deviations, $s_R$ , for the exercises.	The $s_R$ has been on average 9% in the 6 exercises.
Calculate $u(Cref)$	Mean number of participants = 12. $u(Cref) = \frac{s_R}{\sqrt{n}} = \frac{9}{\sqrt{12}} = 2.6 \%$

The bias has been 2 %, 7 %, -2 %, 3 %, 6 % and 5%, in the 6 interlaboratory comparisons where the laboratory has participated.

<b>3</b>	<b><i>Quantify Method and laboratory bias</i></b>	$RMS_{bias} = 4.6 \%$ , $u(Cref) = 2.6 \%$
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<b>4</b>	<b><i>Convert components to standard uncertainty <math>u(x)</math></i></b>	$u(bias) = \sqrt{RMS_{bias}^2 + u(Cref)^2} =$ $= \sqrt{4.6^2 + 2.6^2} = 5.3\%$
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### 5.3 Recovery

Recovery tests, for example the recovery of a standard addition to a sample in the validation process, can be used to estimate the systematic error. In this way, validation data can provide a valuable input to the estimation of the uncertainty.

In an experiment the recoveries for an added spike were 95 %, 98 %, 97 %, 96 %, 99 % and 96 % for 6 **different** sample matrices. The average is 96.8 %. The spike of 0.5 mL was added with a micropipette.

<b>Uncertainty component from the definition of 100% recovery, <math>u(C_{recovery})</math></b>	
<b>Step</b>	<b>Example</b>
Uncertainty of the 100% recovery. Main components are concentration, $u(conc)$ of standard and volume added $u(vol)$	$u(conc)$ - Certificate $\pm 1.2$ % (95 % conf. limit) gives = 0.6 % $u(vol)$ - This value can normally be found in the manufacturer's specifications, or better use the limits specified in your laboratory. Max bias 1 % (rectangular interval) and repeatability max 0.5 % $u(vol) = \sqrt{\left(\frac{1}{\sqrt{3}}\right)^2 + 0.5^2} = 0.76 \%$
Calculate $u(C_{recovery})$	$\sqrt{u(conc)^2 + u(vol)^2} = \sqrt{0.6^2 + 0.76^2} = 1.0 \%$

**3** **Quantify Method and laboratory bias**

$$RMS_{bias} = 3.44 \%$$

$$u(C_{recovery}) = 1.0 \%$$

**4** **Convert components to standard uncertainty  $u(x)$**

$$u(bias) = \sqrt{RMS_{bias}^2 + u(C_{ref})^2} =$$

$$= \sqrt{3.44^2 + 1.0^2} = 3.6 \%$$

## 6 Reproducibility between laboratories, $s_R$

If the demand on uncertainty is low, it can be possible to directly use the  $s_R$  from interlaboratory comparisons as an approximation of  $u_c$  /8/. In such case the expanded uncertainty becomes  $U = 2 \cdot s_R$ . This may be an overestimate depending on the quality of the laboratory – worst-case scenario. It may also be an underestimate due to sample inhomogeneity or matrix variations.

### 6.1 Data given in standard method

In order to use a figure taken directly from the standard method, the laboratory must prove that they are able to perform in accordance with the standard method /8/, i.e. demonstrating control of bias and verification of the repeatability,  $s_r$ . Reproducibility data can either be given as a standard deviation  $s_R$  or as reproducibility limit  $R$  and then  $s_R = R/2.8$

The example below is taken from ISO/DIS 15586 *Water Quality — Determination of trace elements by atomic absorption spectrometry with graphite furnace*. The matrix is wastewater. Combined uncertainty in wastewater,  $u_c$ , is taken from the  $s_R$  from interlaboratory comparison exercises quoted in the ISO method.

**Table 1** ISO/DIS 15586 - Results from the interlaboratory comparison – Cd in water with graphite furnace AAS. The wastewater was digested by the participants.

Cd		n	Outliers	Nominal value μg/L	Mean μg/L	Recovery, %	$s_r$ %	$s_R$ %
Synthetic	Lower	33	1	0.3	0.303	101	3.5	17.0
Synthetic	Higher	34	2	2.7	2.81	104	1.9	10.7
Fresh water	Lower	31	2		0.572		2.9	14.9
Fresh water	Higher	31	3		3.07		2.1	10.4
Waste water		27	2		1.00		3.1	27.5

Measurand	Combined Uncertainty $u_c$	Expanded Uncertainty $U$
Cd	$u_c = 27.5 \%$	$2 \cdot u_c = 55 \% \approx 50 \%$

### 6.2 Data from interlaboratory comparisons

Interlaboratory comparisons are valuable tools in uncertainty evaluation. The reproducibility between laboratories is normally given directly in reports from the exercises as  $s_R$ .

These data may well be used by a laboratory (having performed satisfactorily in the comparisons) as the standard uncertainty of the analysed parameter, provided that the comparison covers all relevant uncertainty components and steps (see /9/,

section 5.4.6.3). For example, a standard deviation in an interlaboratory comparison,  $s_R$ , can be directly used as a combined standard uncertainty,  $u_c$ .

**Table 2** Summary results (mean values) from 10 interlaboratory comparisons that Lab A has participated in. The reproducibility standard deviation is given in absolute units,  $s_R$  and in relative units  $s_R$  %.

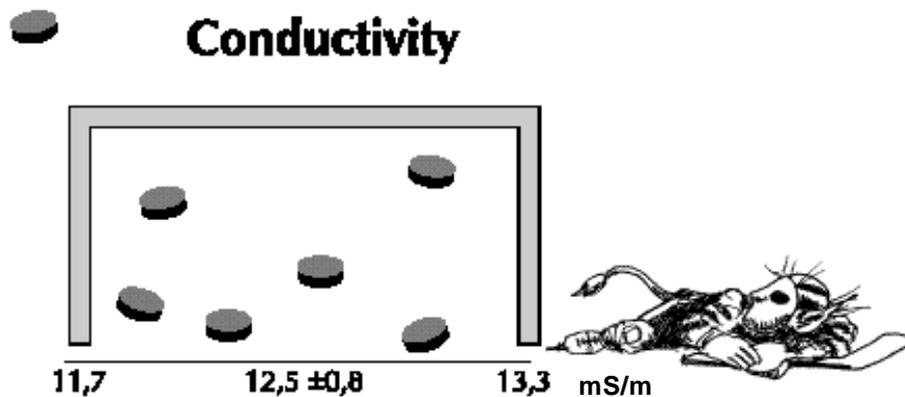
Variable	Nominal value	Lab A % deviation	$s_R$ (abs)	$s_R$ %	No. of labs	Excluded
pH	7.64	-0.037	0.101		90	5
Conductivity, mS/m	12.5	-2.8	0.40	3.2	86	6
Alkalinity, mmol/L	0.673	2.3	0.026	3.9	60	3
Turbidity, FNU	1.4	-9.1	0.1	14.2	44	3
NH <sub>4</sub> -N, µg/L	146	2.2	12.0	8.8	34	5
NO <sub>3</sub> -N, µg/L	432	-1.6	16.3	3.7	39	6

In Table 2 we find that for conductivity, for instance, the mean value for the results from 10 interlaboratory comparisons is 12.5 mS/m. The reproducibility relative standard deviation is 0.4 (3.2 %), which is an average (or pooled) standard deviation between the laboratories in the different interlaboratory comparisons and this value can be taken as an estimate of combined uncertainty i.e.

$$u_c(\text{conductivity}) = s_R = 0.4 \text{ mS/m, thus } U = 2 \cdot 0.4 = 0.8 \text{ mS/m}$$

If we take the ammonium results, we have a mean nominal value of 146 µg/L, and we find that the reproducibility,  $s_R$ , is 8.8 %. Thus  $U = 2 \cdot 8.8 = 17.6 = 18$  % at this concentration level.

Comment: In Section 3 the expanded uncertainty for ammonium is 5 % using an automated method in one highly qualified laboratory.



## 7 Examples

In this chapter, practical examples on how measurement uncertainty can be calculated using the method of this handbook are given.

### 7.1 Ammonium in water

Ammonium in water has already been treated in section 3.2 and section 6.2 . The results are summarised in Table 3.

**Table 3 Measurement uncertainty of ammonium in water – comparison of different calculations**

Uncertainty calculations based on	Relative expanded uncertainty, U	Comment
Control sample + proficiency testing	± 6 %	Uncertainty for one good laboratory- level 200 µg/L.
Interlaboratory comparisons	± 18 %	Uncertainty in general among laboratories – level 150 µg/L

### 7.2 BOD in wastewater

Biological Oxygen Demand, BOD, is a standard parameter in the monitoring of wastewater. This example shows how data from ordinary internal quality control can be used together with CRM results or data from interlaboratory comparison exercises to calculate the within-lab reproducibility and bias components of the measurement uncertainty. The results are summarised in Table 4

**Table 4 Measurement uncertainty of BOD in water - comparison of different calculations**

Uncertainty calculations based on	Relative expanded uncertainty, U	Comment
Control sample + CRM	± 10 %	
Control sample + interlaboratory comparisons	± 10 %	n = 3, unreliable estimate
Interlaboratory comparisons	± 16 %	Uncertainty in general among laboratories

For BOD at high concentrations, using the dilution analytical method, the major error sources are the actual oxygen measurement and variation in the quality of the seeding solution. These errors will be included in the calculations.

The raw data from the internal quality control, using a CRM, used for the calculations is shown in Appendix 7.

The laboratory has only participated in three interlaboratory comparison exercises the last 2 years (Table 5). At least six would be needed, so here we estimate the bias two different ways – with CRM and with interlaboratory comparisons.

**Table 5 BOD - results from interlaboratory comparisons**

<b>Exercise</b>	<b>Nominal value</b>	<b>Laboratory result</b>	<b>Bias</b>	$s_R$	<b>Number of labs</b>
	mg/L	mg/L	%	%	
1	154	161	+ 4.5	7.2	23
2	219	210	- 4.1	6.6	25
3	176	180	+2.3	9.8	19
$\bar{X}$			+0.9	7.87 <sup>3</sup>	22.3
$RMS_{bias}$			3.76	-	-

---

<sup>3</sup> If  $s_R$  or the number of participants vary substantially from exercise to exercise, then a pooled standard deviation will be more correct to use. In this case, where the variation in  $s_R$  is limited, we simply calculate the mean  $s_R$  (the corresponding pooled standard deviation becomes 7.82, an insignificant difference).

**Example A: BOD with Internal quality control + a CRM**

Step	Action	Example: BOD in wastewater
1	Specify Measurand	BOD in wastewater, measured with EN1899-1 (method with dilution, seeding and ATU). The demand on uncertainty is $\pm 20\%$ .
2	Quantify $u(R_w)$  A control sample  B possible steps not covered by the control sample	A: The control sample, which is a CRM, gives an $s = 2.6\%$ at a level of 206 mg/L O <sub>2</sub> . $s = 2.6\%$ is also when setting the control chart limits.  B: The analysis of the control sample includes all analytical steps after sampling
3	Quantify Method and laboratory bias	The CRM is certified to 206 $\pm 5$ mg/L O <sub>2</sub> . The average result of the control chart is 214.8. Thus, there is a bias of 8.8 mg/L = 4.3%.  The $s_{bias}$ is 2.6% (n=19)  The $u(Cref)$ is 5 mg/L / 1.96 = 1.2%
4	Convert components to standard uncertainty $u(x)$	$u(R_w) = 2.6\%$  $u(bias) = \sqrt{bias^2 + \frac{s_{bias}^2}{n} + u(Cref)^2}$ $= \sqrt{4.3^2 + \left(\frac{2.6}{\sqrt{19}}\right)^2 + 1.2^2} = 4.5\%$
5	Calculate combined standard uncertainty, $u_c$	$u_c = \sqrt{2.6^2 + 4.5^2} = 5.2\%$
6	Calculate expanded uncertainty, $U = 2 \cdot u_c$	$U = 2 \cdot 5.2 = 10.4 \approx 10\%$

**Example B: BOD with Internal quality control + interlaboratory comparison results**

Step	Action	Example: BOD in wastewater
1	Specify Measurand	BOD in wastewater, measured with EN1899-1 (method with dilution, seeding and ATU). The demand on uncertainty is $\pm 20\%$ .
2	Quantify $u(R_w)$ A control sample B possible steps not covered by the control sample	A: The control sample, which is a CRM, gives an $s$ of $2.6\%$ at a level of $206\text{ mg/L O}_2$ . $s = 2.6\%$ is also used as $s$ when setting the control chart limits. B: The analysis of the control sample includes all analytical steps after sampling
3	Quantify Method and laboratory bias Data from Table 5	$RMS_{bias} = 3.76$ $u(C_{ref}) = \frac{s_R}{\sqrt{n}} = \frac{7.9}{\sqrt{22.3}} = 1.67$
4	Convert components to standard uncertainty $u(x)$	$u(R_w) = 2.6\%$ $u(bias) = \sqrt{RMS_{bias}^2 + u(C_{ref})^2} = \sqrt{3.76^2 + 1.67^2} = 4.11\%$
5	Calculate combined standard uncertainty, $u_c$	$u_c = \sqrt{2.6^2 + 4.11^2} = 4.86\%$
6	Calculate expanded uncertainty, $U = 2 \cdot u_c$	$U = 2 \cdot 4.86 = 9.7 \approx 10\%$

### 7.3 PCB in sediment

In this example, the  $u(R_w)$  is estimated from a quality control sample and the  $u(\text{bias})$  is estimated from two different sources: in the first example the use of a CRM and in the second example participation in interlaboratory comparisons. In the summary table both ways of calculating the  $u(\text{bias})$  will be compared.

For this analysis, the sample-work up is a major error source (both for random and systematic errors), and it is thus crucial that this step is included in the calculations. The number of interlaboratory comparisons is too few to get a good estimate.

#### Example C: PCB with Internal quality control + a CRM

Step	Action	Example: PCB in sediment
1	Specify Measurand	Sum of 7 PCB:s in sediment by extraction and GC-MS(SIM). Demand on expanded uncertainty is $\pm 20 \%$ .
2	Quantify $u(R_w)$ A control sample B possible steps not covered by the control sample	A: The control sample, which is a CRM, gives an $s_{R_w} = 8 \%$ at a level of $150 \mu\text{g/kg}$ dry matter. $s_{R_w} = 8 \%$ is also used when setting the control chart limits. B: The analysis of the control sample includes all steps except for drying the sample to determine the dry weight. The uncertainty contribution from that step is considered small and is not accounted for.
3	Quantify method and laboratory bias	The CRM is certified to $152 \pm 14 \mu\text{g/kg}$ . The average result of the control chart is 144. Thus, there is a bias = $5.3 \%$ . The $s_{\text{bias}} = 8 \%$ (n=22) $u(\text{Cref})$ $14 \mu\text{g/kg}/1.96$ , which is $4.7 \%$ relative.
4	Convert components to standard uncertainty $u(x)$	$u(R_w) = 8 \%$ $u(\text{bias}) = \sqrt{\text{bias}^2 + \frac{s_{\text{bias}}^2}{\sqrt{n}} + u(\text{Cref})^2}$ $= \sqrt{5.3^2 + \left(\frac{8}{\sqrt{22}}\right)^2 + 4.7^2} = 7.29$
5	Calculate combined standard uncertainty, $u_c$	$u_c = \sqrt{8^2 + 7.29^2} = 10.8 \%$
6	Calculate expanded uncertainty, $U = 2 \cdot u_c$	$U = 2 \cdot 10.8 = 21.6 \approx 22 \%$

**Example D: PCB with Internal quality control + interlaboratory comparison**

Step	Action	Example: PCB in sediment
1	Specify Measurand	Sum of 7 PCB:s in sediment by extraction and GC-MS(SIM). Demand on expanded uncertainty is 20 %.
2	Quantify $u(R_w)$ A control sample B possible steps not covered by the control sample	A: The control sample, which is a stable in-house material, gives $s_{Rw} = 8\%$ at a level of 150 $\mu\text{g/kg}$ dry matter. $s_{Rw} = 8\%$ is also used as $s$ when setting the control chart limits. B: The analysis of the control sample includes all steps except for drying the sample to determine the dry weight. The uncertainty contribution from that step is considered small and is not accounted for.
3	Quantify Method and laboratory bias	Participation in 3 interlaboratory comparisons with concentration levels similar to the internal quality control. The bias in the 3 exercises has been $-2\%$ , $-12\%$ and $-5\%$ . $RMS_{bias} = 7.6$ The $s_R$ in the three exercises has been 12 %, 10 % and 11 %, on average $s_R = 11\%$ ( $n=14$ ) $u(Cref) = \frac{11}{\sqrt{14}} = 2.9\%$
4	Convert components to standard uncertainty $u(x)$	The $u(R_w)$ is 8 % $u(bias) = \sqrt{RMS_{bias}^2 + u(Cref)^2} = \sqrt{7.6^2 + 2.9^2} = 8.1\%$
5	Calculate combined standard uncertainty, $u_c$	$u_c = \sqrt{8^2 + 8.1^2} = 11.4$
6	Calculate expanded uncertainty, $U = 2 \cdot u_c$	$U = 2 \cdot 11.4 = 22.8 \approx 23\%$

## Summary table for PCB measurement uncertainty calculations

### *PCB in sediment by extraction and GC-MS (SIM)*

Measurement uncertainty  $U$  (95 % confidence interval) is estimated to  $\pm 20$  % (relative) for 7 PCB:s in sediments at a level of 150  $\mu\text{g}/\text{kg}$  dry weight. The customer demand is  $\pm 20$  %. The calculations are based on internal quality control using a stable sample, CRM and the participation in a limited amount of interlaboratory comparison exercises.

		<b>Value</b>	<b><math>u(x)</math></b>	<b>Comments</b>
<b>Reproducibility within-laboratory, <math>R_w</math></b>				
Control sample $\bar{X} = 160 \mu\text{g}/\text{kg}$ dry weight	$u(R_w)$	12.8 $\mu\text{g}/\text{kg}$ dry weight	8 %	
Other components		too small to be considered		
<b>Method and laboratory bias</b>				
<b>Reference material</b>		Bias: 5.3 % $s_{bias} = 8$ ; $n = 22$ $u(Cref) = 4,7$ %	$u(bias) = 5.88$	$u(bias) = \sqrt{bias^2 + \frac{s_{bias}^2}{n} + u(Cref)^2}$
<b>Interlaboratory comparison</b> $n = 3$		$RMS_{bias} = 7.6$ $u(Cref) = 2.9$ %	$u(bias) = 8.1$	$u(bias) = \sqrt{RMS_{bias}^2 + u(Cref)^2}$

Combined uncertainty,  $u_c$ , is calculated from internal quality control and the maximum bias - interlaboratory comparisons.

<b>Measurand</b>	<b>Combined Uncertainty <math>u_c</math></b>	<b>Expanded Uncertainty <math>U</math></b>
PCB	$u_c = \sqrt{8^2 + 8.1^2} = 11.4$	$U = 2 \cdot u_c = 2 \cdot 11.4 = 22.8 \approx 23$ %

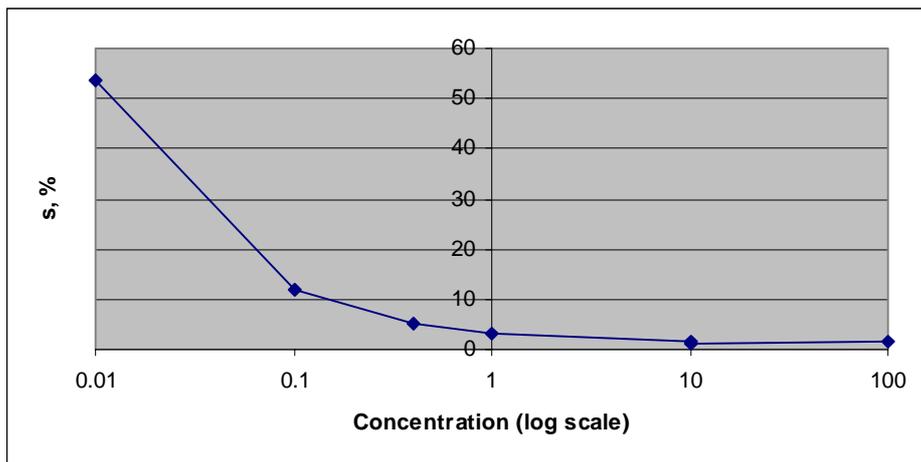
**Conclusion:** In this case the calculation of the  $u(bias)$  gives similar results regardless of whether CRM or interlaboratory comparison results are used. Sometimes interlaboratory comparisons will give considerably higher values, and it might in such cases be more correct to use the CRM results.

## 7.4 Concentration ranges

The measurement uncertainty will normally vary with concentration, both in absolute and relative terms. If the concentration range of the reported data is large, it is thus often necessary to take this into account. For lead (Pb) in water, a recovery experiment was carried out a number of times to investigate within-lab reproducibility over the measurable range – the major component of the measurement uncertainty at low levels. The following results were obtained:

**Table 6 Within-lab reproducibility and recovery for Pb determined with ICP-MS at different concentration levels.**

Addition, $\mu\text{g/L}$	Pb, % recovery	s, %
0.01	109.7	53.8
0.1	125.2	12.1
0.4	91.8	5
1	98.4	3.0
10	98	1.7
10	100.5	1.3
100	105.5	1.4



**Figure 4 Within-lab reproducibility for Pb over the concentration range**

It is clear from the results that the measurement uncertainty, here represented by  $s$ , is strongly concentration dependent. Two approaches are recommended for using these data:

- (1) To divide the measurable range into several parts, and use a fixed relative measurement uncertainty or absolute uncertainty – see Table 7.

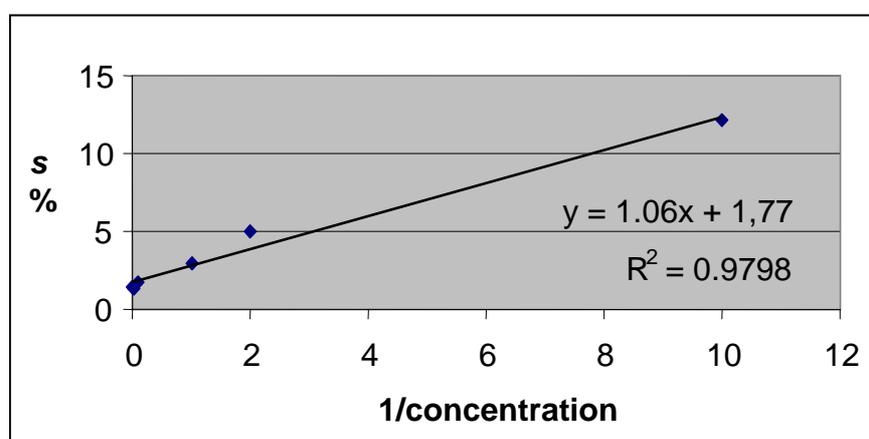
**Table 7 Within-lab reproducibility for Pb divided into three concentration ranges**

Within-lab reproducibility Pb		
Range (µg/L)	<i>s</i> (rel)	<i>s</i> (rel) or (abs)
< 0.1	50 %	0.01 (µg/L)
0.1 - 10	10 %	10 %
> 10	2 %	2 %

In the second column *s* is relative and given in %. In the third column *s* is also relative but an absolute value is given in the lower range close to the detection limit.

(2) To use an equation that describes how the measurement uncertainty varies with concentration

Plotting *s* % against 1/concentration gives a straight line, and a relatively simple equation. (see Figure 5).



**Figure 5:** The relationship between within-lab reproducibility and the inverted concentration for Pb in the range 0.1 – 100 µg/L.

The straight-line equation above tells us that the within-lab reproducibility equals 1.06 multiplied with 1/concentration plus 1.77. For example, at a concentration of 2 µg/L the within-lab reproducibility becomes  $1.06 \cdot 1/2 + 1.77 = 2.3$  %. When reporting to customers, the laboratory can choose between quoting the formula or calculating the measurement uncertainty for each value, using the formula. For further reading, see for example /2/.

## 8 Reporting uncertainty

This is an example on what a data report could look like, when measurement uncertainty has been calculated and is reported together with the data. The company and accreditation body logotypes are omitted, and the report does not contain all information normally required for an accredited laboratory. It is recommended to use either relative or absolute values for the benefit of the customer.

### Analytical Report

Sample identification: P1 – P4

Samples received: 14 December 2002

Analysis period: 14 –16 December 2002

#### Results

##### **NH<sub>4</sub>-N (µg/L):**

<u>Sample</u>	<u>Result</u>	<u>U</u>	<u>Method</u>
P1	103	±6%	23B
P2	122	±6%	23B
P3	12	±10%	23B
P4	14	±10%	23B

##### **TOC (mg/L)**

<u>Sample</u>	<u>Result</u>	<u>U</u>	<u>Method</u>
P1	40	±4.0	12-3
P2	35	±3.5	12-3
P3	10	±1.0	12-3
P4	9	±0.9	12-3

*Signed: Dr Analyst*

The laboratory should also prepare a note explaining how the measurement uncertainty has been calculated for the different parameters. Normally, such an explanatory note should be communicated to regular customers and other customers who ask for information. An example is given below:

**Note on measurement uncertainty from Dr Analyst's laboratory**

**Measurement uncertainty:**

*U* = expanded Measurement Uncertainty, estimated from control sample results, interlaboratory comparison and the analyses of CRMs, using a coverage factor of 2 to reach approximately 95% confidence level.

**NH<sub>4</sub>-N:** *U* is estimated to 6% above 100 µg/L and 10% below 100 µg/L.

**TOC:** *U* is estimated to 10% over the whole concentration range.

**References:**

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EURACHEM/CITAC Guide
- Handbook for calculation of measurement uncertainty in environmental laboratories

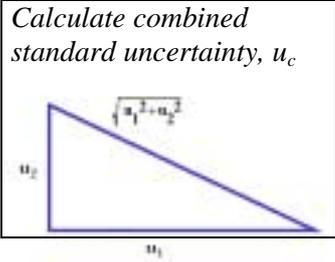
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# 10 Appendices

## Appendix 1: Empty flow scheme for calculations

**Before starting:** Always identify the main error sources, to make sure that they are included in the calculations.

Step	Action	Measurand:
1	Specify Measurand	(measurand) in (matrix) by (method) The customer demand on expanded uncertainty is $\pm$ _ %.
2	Quantify $u(R_w)$ A control sample B possible steps not covered by the control sample	A:  B:
3	Quantify method and laboratory bias	
4	Convert components to standard uncertainty $u(x)$	
5	Calculate combined standard uncertainty, $u_c$ 	
6	Calculate expanded uncertainty, $U = 2 \cdot u_c$	

## Appendix 2: Empty summary table

(*measurand*) in (*matrix*) by (*method*)

Measurement uncertainty  $U$  (95 % confidence interval) is estimated to  $\pm$  \_ % (relative) for (*measurand*) in (*matrix*) at a level of \_ (unit). The customer demand is  $\pm$  \_ %. The calculations are based on (control samples/control limits/CRM/interlaboratory comparison/other).

		<i>Value</i>	<i>Relative <math>u(x)</math></i>	<i>Comments</i>
<b>Reproducibility within-laboratory, <math>R_w</math></b>				
Control sample $\bar{X} = (\text{conc}) (\text{unit})$	$s_{Rw}$			
Other components				
<b>Method and laboratory bias</b>				
Reference material	bias			
Interlaboratory comparison	bias			
Recovery test	bias			
<b>Reproducibility between laboratories</b>				
Interlaboratory comparison	$s_R$			
Standard method	$s_R$			

Combined uncertainty,  $u_c$ , is calculated from \_\_\_ and bias from \_\_\_.

<i>Measurand</i>	<i>Combined Uncertainty <math>u_c</math></i>	<i>Expanded Uncertainty <math>U</math></i>
		$2 \cdot u_c =$

### Appendix 3: Error model used in this handbook

This model is a simplification of the model presented in the ISO guide /8/:

$$y = m + (\delta + B) + e$$

$y$  measurement result of a sample

$m$  expected value for  $y$

$\delta$  method bias

$B$  laboratory bias – the uncertainty for these are combined to  $u(bias)$

$e$  random error at within-laboratory reproducibility conditions,  $R_w$

#### Uncertainty estimation in section 3 to 5

$$u(y)^2 = s_{Rw}^2 + u(bias)^2$$

$s_{Rw}^2$	The estimated variance of $e$ under within-laboratory reproducibility conditions – intermediate precision. In the ISO guide the repeatability, $s_r$ is used as an estimate of $e$ .
$u(bias)^2$	The estimated variance of method bias and laboratory bias.

#### Uncertainty estimation in section 6

The combined uncertainty  $u(y)$  or  $u_c$  can also be estimated by from reproducibility data.

$$u(y)^2 = s_L^2 + s_r^2 = s_R^2 - \text{equation A6 ref. /8/}$$

where  $s_R^2$  is the estimated variance under reproducibility conditions and where  $s_L^2$  is either the estimated variance of  $B$  if one method is used by all laboratories or an estimated variance of  $B$  and  $\delta$  if several different methods have been used in the collaborative study and  $s_r^2$  is the estimated variance of  $e$ .

#### Comment

For samples that are more inhomogeneous and have big variations in matrix the estimation of the measurement uncertainty of the method can become too low. However we recommend the use of repeatability limit for duplicate analyses  $r = 2.8 \cdot s_r$  in order to control sample inhomogeneity.

## Appendix 4: Uncertainty of bias for NH<sub>4</sub>-N in section 3.2

Results for a laboratory from interlaboratory comparisons of NH<sub>4</sub>-N in water.

Exercise	Nominal value $x_{ref}$	Laboratory result $x_i$	Bias	$s_R$	Number of labs
	mg/L	mg/L	%	%	
1999 1	81	83	2.4	10	31
2	73	75	2.7	7	36
2000 1	264	269	1.9	8	32
2	210	213	1.4	10	35
2001 1	110	112	1.8	7	36
2	140	144	2.9	11	34
$\bar{X}$			<b>+ 2.18</b>	8.8	34
<i>RMS</i>			<b>2.25</b>	-	-

$$RMS \text{ of the bias} = \sqrt{\frac{\sum bias_i^2}{n}} = \sqrt{\frac{2.4^2 + 2.7^2 + \dots + 2.9^2}{n}} = 2.25 \% \text{ (rel)}$$

$$u(C_{ref}) = \frac{s_R}{\sqrt{n}} = \frac{8.8}{\sqrt{34}} = 1.5 \% \text{ (rel)}$$

A t-test shows that the bias (+2.18 %) is not significant (t = 0.6). However, in order not to complicate the calculations when the bias is small, t-test are normally not performed.

The mean value of  $s_R$  is used. If differences in number of laboratories and  $s_R$  are very big pooled standard deviations should be used. In this case the pooled standard deviation is 8.9 % for  $s_R$  which is the same as the mean value of 8.8 %.

## Appendix 5: Raw data for NH<sub>4</sub>-N in section 4.3

The estimation of the standard deviation from the range is explained in Appendix 8

concentration < 15 µg/L

Sample	X1	X2	$\bar{X} = \frac{x_{i1} + x_{i2}}{2}$	$d = x_{i1} - x_{i2}$	$100 \cdot \frac{ d }{\bar{X}} = r\%$
1	7.46	7.25	7.355	0.210	2.855
2	9.01	9.17	9.090	-0.160	1.760
3	3.6	3.1	3.350	0.500	14.925
4	6.48	6.48	6.480	0.000	0.000
5	14.49	14.12	14.305	0.370	2.587
6	10.84	9.89	10.365	0.950	9.165
7	4.61	5	4.805	-0.390	8.117
8	2.6	2.42	2.510	0.180	7.171
9	2.8	2.62	2.710	0.180	6.642
10	5.84	6.19	6.015	-0.350	5.819
11	2.12	2.5	2.310	-0.380	16.450
12	2.3	2.11	2.205	0.190	8.617
13	2.52	2.89	2.705	-0.370	13.678
14	3.71	3.71	3.710	0.000	0.000
15	7.43	7.43	7.430	0.000	0.000
16	8.83	8.51	8.670	0.320	3.691
17	9.12	8.79	8.955	0.330	3.685
18	8.24	7.9	8.070	0.340	4.213
19	2.62	2.78	2.700	-0.160	5.926
20	3.33	3.33	3.330	0.000	0.000
21	2.69	2.69	2.690	0.000	0.000
22	12.09	12.09	12.090	0.000	0.000
23	4.24	4.24	4.240	0.000	0.000
24	10.49	10.64	10.565	-0.150	1.420
25	3.68	3.52	3.600	0.160	4.444
26	9.37	9.37	9.370	0.000	0.000
27	2.22	2.06	2.140	0.160	7.477
28	6.1	6.1	6.100	0.000	0.000
29	2.96	2.86	2.910	0.100	3.436
30	14.02	13.7	13.860	0.320	2.309
31	4.24	3.62	3.930	0.620	15.776
32	5.1	4.61	4.855	0.490	10.093
33	2.78	2.62	2.700	0.160	5.926
34	8.52	6.81	7.665	1.710	22.309
35	12.82	14.05	13.435	-1.230	9.155
36	3.17	2.4	2.785	0.770	27.648
37	11.28	11.43	11.355	-0.150	1.321
38	14.31	13.82	14.065	0.490	3.484
39	4.01	4.48	4.245	-0.470	11.072
40	3.27	3.58	3.425	-0.310	9.051
41	9.98	10.29	10.135	-0.310	3.059
42	12.56	13.66	13.110	-1.100	8.391
43	3.35	2.88	3.115	0.470	15.088
<b>Mean:</b>			<b>6.499</b>		<b>6.4363</b>

= mean range (%)

**s(r) % = range(mean)/1.128 = 5.71 %**

concentration > 15 µg/L

Sample	X1	X2	$\bar{X} = \frac{x_{i1} + x_{i2}}{2}$	$d = x_{i1} - x_{i2}$	$100 \cdot \frac{ d }{\bar{X}} = r\%$
1	37.62	36.85	37.235	0.770	2.068
2	16.18	16.56	16.370	-0.380	2.321
3	28.82	28.65	28.735	0.170	0.592
4	4490	4413	4451.500	77.000	1.730
5	135.7	124.7	130.200	11.000	8.449
6	62.56	62.25	62.405	0.310	0.497
7	158.9	159.2	159.050	-0.300	0.189
8	16540	16080	16310.000	460.000	2.820
9	31.26	30.12	30.690	1.140	3.715
10	58.49	60.11	59.300	-1.620	2.732
11	740.5	796.2	768.350	-55.700	7.249
12	130.3	126.9	128.600	3.400	2.644
13	29.35	29.19	29.270	0.160	0.547
14	1372	1388	1380.000	-16.000	1.159
15	36.55	44.74	40.645	-8.190	20.150
16	22.57	23.37	22.970	-0.800	3.483
17	34.75	33.15	33.950	1.600	4.713
18	92.93	94.01	93.470	-1.080	1.155
19	40.6	42.23	41.415	-1.630	3.936
20	80.36	86.36	83.360	-6.000	7.198
21	15.76	18.54	17.150	-2.780	16.210
22	78.22	73.76	75.990	4.460	5.869
23	48.89	50.91	49.900	-2.020	4.048
24	17.65	16.72	17.185	0.930	5.412
25	36.56	35.3	35.930	1.260	3.507
26	51.89	52.2	52.045	-0.310	0.596
27	197.5	206.5	202.000	-9.000	4.455
28	70.32	69.22	69.770	1.100	1.577
29	29.99	30.62	30.305	-0.630	2.079
30	31.9	32.36	32.130	-0.460	1.432
<b>Mean:</b>			<b>816.331</b>		<b>4.0843</b> = mean range (%)
			<b>s(r) % = range(mean)/1.128 =</b>		<b>3.62</b> %

## Appendix 6: Raw data for oxygen in Section 4.4

Data plotted in Figure 3. "Range" equals the absolute value of the difference between Result 1 and Result 2.

Res. 1 m g /L	Res. 2 m g /L	Range m g /L
8.90	8.91	0.01
8.99	9.01	0.02
8.90	8.90	0.00
9.11	9.12	0.01
8.68	8.64	0.04
8.60	8.51	0.09
8.81	8.81	0.00
8.02	8.00	0.02
7.05	7.08	0.03
6.98	7.01	0.03
7.13	7.16	0.03
6.79	6.78	0.01
6.55	6.53	0.02
4.68	4.68	0.00
5.28	5.33	0.05
7.42	7.40	0.02
7.62	7.63	0.01
5.88	5.88	0.00
6.03	6.06	0.03
6.33	6.33	0.00
5.90	5.90	0.00
6.24	6.27	0.03
6.02	6.02	0.00
9.13	9.11	0.02
9.10	9.14	0.04
8.50	8.44	0.06
8.73	8.71	0.02
8.09	8.09	0.00
7.56	7.58	0.02
6.30	6.32	0.02
6.43	6.44	0.01
7.25	7.34	0.09
7.28	7.31	0.03
8.00	8.03	0.03
8.38	8.29	0.09
9.23	9.29	0.06
9.09	9.08	0.01
9.37	9.36	0.01
9.38	9.37	0.01
9.32	9.25	0.07
8.47	8.49	0.02
8.27	8.28	0.01
8.37	8.31	0.06
8.09	8.15	0.06
8.05	8.03	0.02
7.38	7.40	0.02
7.49	7.49	0.00
4.52	4.49	0.03
4.45	4.44	0.01
4.29	4.27	0.02
mean range :		0.026
mean range / 1.128 :		0.024

## Appendix 7: Raw data for BOD in example A and B

Results in mg/L O<sub>2</sub> consumption. The certified value of the CRM is 206 ± 5 mg/L. As the average of two results is always reported for ordinary samples, the s is also calculated from the average of each sample pair in the internal quality control.

Date	Res. 1	Res. 2	Average
12-09-00	218.90	214.77	216.84
01-03-01	206.46	220.83	213.65
13-03-01	221.18	210.18	215.68
02-04-01	215.00	206.50	210.75
14-08-01	194.96	218.03	206.50
05-09-01	218.65	216.55	217.60
19-09-01	223.86	212.19	218.03
16-10-01	215.58	213.01	214.30
07-11-01	196.26	214.93	205.60
28-11-01	210.89	206.89	208.89
11-12-01	228.40	222.73	225.57
13-12-01	206.73	229.03	217.88
15-01-02	207.00	208.47	207.74
22-01-02	224.49	213.66	219.08
30-01-02	201.09	214.07	207.58
11-02-02	218.83	223.13	220.98
06-03-02	216.69	218.22	217.46
18-09-02	206.36	227.96	217.16
02-10-02	215.21	226.18	220.70
Average:			214.84
s:			5.58
s%:			2.60

## Appendix 8: Estimation of standard deviation from range

Number of samples	Factor , $d_2$	<p>Estimation of standard deviation from range (max-min), /1/ and /13, page 11/.</p> <p>The standard deviation, <math>s</math> can be estimated from</p> $s = \frac{\text{range}}{d_2}$
n=2	1.128	
n=3	1.693	
n=4	2.059	
n=5	2.326	
n=6	2.534	
n=7	2.704	
n=8	2.847	
n=9	2.970	
n=10	3.078	
<b>For comparison</b>		
Rectangular interval	3.464	(Example, see Appendix 5 and 6)
95 % conf. limit.	3.92	



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